It is furthermore preferred that the filter is a filter membrane which is optionally or preferably contained in a microtitre plate. Additionally preferred is the use of SDS as detergent or TRITON X-100™ for non-β-amyloid aggregates.

On page 25, fourth paragraph, fifth line, delete "Triton X-100" and replace with --TRITON X-100™--.

Cells were washed with buffer A [50 mM sodium phosphate (pH 8), 150 mM NaCl, and 1 mM EDTA]. If necessary, the cell pallet was stored at -70°C. Cells were resuspended in 25 ml buffer A. PMSF and lysozyme (Boehringer Mannheim) were added to 1 mM and 0.5 mg/ml, respectively, and incubated on ice for 45 min. Cells were lysed by sonication (2 x 45 s, 1 min cooling, 200-300 Watt), and TRITON X-100™ was added to a final concentration of 0.1% (v/v). The lysate was centrifuged at 30.000 x g for 30 min, and the supernatant was collected.

On page 25, paragraph five, line 4, delete "Triton X-100" and replace with --TRITON X-100™--.

5 ml of a 1:1 slurry of GST-agarose (Sigma), previously equilibrated in buffer A, was added and the mixture was stirred for 30 min. The slurry was poured into a 1.6 cm diameter column, washed once with 40 ml buffer A containing 1 mM PMSF and 0.1% TRITON X-100™ and twice with 40 ml buffer A containing 1 mM PMSF. The protein was eluted with 5 x 2 ml buffer A containing 15 mM reduced glutathione (Sigma). Aliquots of the fractions were analyzed by SDS-PAGE and the fractions containing purified GST fusion protein were combined. Finally, the pooled fractions were dialysed.

IN THE CLAIMS

Please amend claims 1, 5, 6, 8-13, 15-21, and 23-25 to read as follows:

1. (Amended) A method of detecting the presence of detergent- or urea-insoluble amyloid-like fibrils or protein aggregates on a filter comprising the following steps: